

Neuroprotective Effects of Herbal Extract (*Rosa canina*, *Tanacetum vulgare* and *Urtica dioica*) on Rat Model of Sporadic Alzheimer's Disease

Parvaneh Daneshmand¹, Kiomars Saliminejad², Marzieh Dehghan Shasaltaneh³, Koorosh Kamali², Gholam Hossein Riazi³, Reza Nazari³, Pedram Azimzadeh⁴, and Hamid Reza Khorram Khorshid^{1*}

1. Genetic Research Center, University of Social Welfare and Rehabilitation Sciences, Tehran, Iran

2. Reproductive Biotechnology Research Center, Avicenna Research Institute, ACECR, Tehran, Iran

3. Laboratory of Neuro-organic Chemistry, Institute of Biochemistry and Biophysics (IBB), University of Tehran, Tehran, Iran

4. Basic and Molecular Epidemiology of Gastrointestinal Disorders Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Abstract

Background: Sporadic Alzheimer's Disease (SAD) is caused by genetic risk factors, aging and oxidative stresses. The herbal extract of *Rosa canina* (*R. canina*), *Tanacetum vulgare* (*T. vulgare*) and *Urtica dioica* (*U. dioica*) has a beneficial role in aging, as an anti-inflammatory and anti-oxidative agent. In this study, the neuroprotective effects of this herbal extract in the rat model of SAD was investigated.

Methods: The rats were divided into control, sham, model, herbal extract -treated and ethanol-treated groups. Drug interventions were started on the 21st day after modeling and each treatment group was given the drugs by intraperitoneal (I.P.) route for 21 days. The expression levels of the five important genes for pathogenesis of SAD including *Syp*, *Psen1*, *Mapk3*, *Map2* and *Tnf- α* were measured by qPCR between the hippocampi of SAD model which were treated by this herbal extract and control groups. The Morris Water Maze was adapted to test spatial learning and memory ability of the rats.

Results: Treatment of the rat model of SAD with herbal extract induced a significant change in expression of *Syp* ($p=0.001$) and *Psen1* ($p=0.029$). In Morris Water Maze, significant changes in spatial learning seen in the rat model group were improved in herbal-treated group.

Conclusion: This herbal extract could have anti-dementia properties and improve spatial learning and memory in SAD rat model.

Avicenna J Med Biotech 2016; 8(3): 120-125

Keywords: Alzheimer disease, Gene expression, Herbal extract

Introduction

Sporadic Alzheimer's Disease (SAD) is a chronic neurodegenerative disorder which is characterized by progressive cognitive impairment, memory loss, and behavioral disturbances¹. SAD, the most controllable type of Alzheimer's disease (AD) by drug intervention, is a multifactorial disease and affected by genetic risk factors, aging and oxidative stresses^{2,3}. The impairment of memory and cognition in AD patients is caused by synaptic loss, enhanced inflammatory signaling, the progressive deposition of senile plaques, neurofibrillary tangles and neurodegeneration⁴⁻⁶.

Synapses are believed to be the basis of AD pathology and synaptophysin (*SYP*) is one of the best targets that is often measured to quantify synapses function. *SYP* mRNA level is reduced in the post-mortem AD brain⁷⁻¹⁰. The amyloid- β (A β) peptides are implicated

in the pathogenesis of AD and derived from abnormal processing of amyloid- β precursor protein (APP) by γ -secretase. Presenilin1 (*PSEN1*) is the catalytic subunit of γ -secretase. Neuronal inflammation and oxidative stresses activate *Psen1* gene expression leading to production of A β plaque and synaptic dysfunction and the effect could be enhanced by hypoxia¹¹. Thus, abnormalities of tau and other neuron cytoskeletal proteins are correlative to the pathogenesis of AD. Among these genes, alteration in expression level of Microtubule-associated protein 2 (*Map2*) which has a vital role in nutrition and plasticity of the neuron was most obvious¹². On the other hand, phosphorylation of the signaling proteins also adjusts neuronal plasticity and APP processing. Among protein kinases that play such role, *Mapk3* has shown significant change of expression

* Corresponding author:
Hamid Reza Khorram Khorshid
MD., Ph.D., Genetic Research
Center, University of Social
Welfare and Rehabilitation
Sciences, Tehran, Iran
Tel/Fax: +98 21 22180138
E-mail:
hrkk1@uswr.ac.ir
Received: 15 Dec 2015
Accepted: 3 Feb 2016

level in AD brain and has a vital role in neurogenesis¹³.

Accumulating evidences have shown that inflammatory reaction induced by A β results in TNF increase, oxidative stresses and memory decline. Up regulation of TNF induced by A β and increased levels of TNF in the brain of Alzheimer's patient have been shown in another study¹⁴. Brain oxidative stresses have been deeply associated with cognitive impairment and Alzheimer's disease progression¹⁵. Enhanced oxidative stress leads to deposition of senile plaque and synaptic loss that result in neurodegeneration. Therefore, one of the most important factors for preventing or treatment of this process is controlling of oxidative stresses. Recently, studies on novel preparation of ethanolic herbal extract from *Rosa canina* (*R. canina*), *Tanacetum vulgare* (*T. vulgare*) and *Urtica dioica* (*U. dioica*) with an immune system modulator effect, have shown positive effects on reduction of oxidative stresses and pro-inflammatory status and they seem to act as anti-aging drugs¹⁶⁻²⁰.

With respect to beneficial effects of this compound in aging, as an anti-inflammatory and anti-oxidative agent, the neuroprotective effects of the her-bal extract by comparing the expression of the five important genes for pathogenesis of AD, *Syp*, *Psen1*, *Mapk3*, *Map2* and *Tnf-a* between the hippocampus of SAD model treated by herbal extract and control groups were investigated. Additionally, therapeutic effects of herbal extract at behavioral, learning and memory levels were studied as well.

Materials and Methods

Experimental animals and grouping

In this study, 40 adult male Wistar rats, weighing 250-300 g (obtained from Pasteur Institute of Iran), were housed two pair per cage with open access to food and water. They were kept in a constant environment at 22°C and 12 hr light/dark cycle. All behavioral experiments were carried out between 11 am to 4 pm.

After one week of housing, rats were randomly divided into five groups, each consisting of eight animals; the control group that received no medication and no surgery, the model group (STZ) which received bilateral Intracerebroventricular (ICV) injection of streptozocin (STZ, purchased from Sigma Chemical Co, St. Louise, USA), five days after surgery, at dose of 3 mg/kg²¹, the sham-operation group (S) that received bilateral ICV injection of a-CSF, as the vehicle of STZ, the herbal extract-treated STZ group (Rose Pharmed Biomedical Inc.), which received compound 21 days after modeling²¹, as I.P. at the dose of 20 mg/kg/day for three weeks²⁰, and the ethanol-treated STZ group (86% ethanol as I.P.) that received alcohol as the vehicle of herbal extract.

After three weeks of treatment, all groups of rats were tested for learning and memory using Morris Water Maze (MWM) test^{22,23}. The rats were sacrificed

after MWM test and the hippocampi were immediately dissected and stored in RNA-Protector at -20°C for later RNA extraction. All experiments were conducted in accordance with the National Institute of Health Guide for the care and use of laboratory animals²⁴.

Surgical procedure

Based on bregma for stereotaxic surgery²¹, the rats were first anesthetized with an intraperitoneal combination of ketamine (50 mg/kg) and xylazine (10 mg/kg) and then their heads were fixed in a stereotaxic apparatus. The scalp was disinfected and incised on the median sagittal of skull and the periosteum was separated. Two small holes were drilled through the skull and injection cannula was lowered into the lateral ventricles (anterior-posterior=-0.9 mm; medial-lateral=1.5 mm; and dorsal-ventral=-3.5 mm). Using a Hamilton syringe, STZ was injected 3 mg/kg and the injection was repeated on the third day with the same dose²⁴. The bilateral ICV injection of STZ was carried out on all groups, except for the control and sham-operated group.

Morris water maze behavioral test

Spatial learning and memory were evaluated using water maze test. The water maze consisted of a circular black tank (140 cm diameter, 60 cm height) which was filled with water (23±1°C) to a depth of 40 cm. Four equal spaces located at the periphery of the tank divided the pool into four quadrants and were used as the start position. An escape platform (11 cm diameter) was set 2 cm under the water at a constant location in the north-east quadrant of the tank. The rats were participated to a daily session of four training trials for five consecutive days²⁴.

The animals were subjected to a daily session of four training trials for five consecutive days, so the rat was permitted to find the platform at maximum time of 60 s and after that it was allowed to remain there for 30 s. If the rat was unable to find the platform within 60 s, it was guided to the platform by the experimenter. The amount of time spent to find the platform [Escape Latency (s)], the distance animal swam before finding it [Path Length (cm)] and the swimming speed [velocity (cm/s)] were measured.

One day after acquisition, a probe test was performed by removing the hidden platform. Rat was allowed to swim in the pool for 60 s. The times spent in the target (zone 3) and opposite (zone 2) quadrants were recorded.

To assess inability of rats to find the hidden platform, they were allowed to the pool for 60 s and four trials on sixth day. A number of behavior tests were carried out using a computer-based video tracking system^{22,23}.

Total RNA extraction and cDNA synthesis

Total RNA was isolated from the hippocampus tissues using RNeasy Plus Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions.

Table 1. Primer sequences and amplicon sizes

Gene		Primer sequence (5'→3')	Product size (bp)
<i>Syp</i>	forward	GGGCCTATGATGGACTTTCT	135
	reverse	TGGGCATCTCCTTGATAATG	
<i>Psen1</i>	forward	CCATATTGATCGGCCGTGTG	140
	reverse	GAAGGGCTGCACGAGATAAT	
<i>TNF-α</i>	forward	CTGAACTTCGGGGTGATCG	152
	reverse	GCTTGGTGGTTTGCTACGAC	
<i>Mapk3</i>	forward	CTGGAAGCCATGAGAGATGTT	203
	reverse	TCGCAGGTGGTGTGATAAG	
<i>Map2</i>	forward	TAAGCGGAAAACCACAGCAAC	131
	reverse	AGGAAGGTCTTGGGAGGGAA	
<i>Actb</i>	forward	ACAACCTTCTTGCAGCTCCTC	199
	reverse	TGACCCATACCCACCATCAC	

The RNA purity and integrity were evaluated using Nano Drop ND-2000 spectrophotometer (Thermo Fisher scientific, Wilmington, USA) and gel electrophoresis.

Next, 1 μg of RNA was used to prepare cDNA with the Revert Aid First Strand cDNA Synthesis kit (Fermentas, Thermo Fisher Scientific) according to the manufacturer's protocol.

Real-time qPCR

The relative expression levels of the five genes *Syp*, *Psen1*, *Mapk3*, *Mtap2* and *Tnf-α* in hippocampus of rats in each group were detected using SYBR green real-time PCR. The real-time qPCR was performed on an ABI 7500 real-time PCR system (Applied Biosystems, Foster City, CA, USA) using Takara SYBR Master Mix instructions (Shiga, Japan). Primers were designed using Genscript and Primer3 online programs.

Primer sequences and amplicon sizes are shown in table 1.

The expression of all target genes were normalized with the expression of *Actb* as the endogenous control^{25,26}. Cycle threshold (Ct) values were used to calculate fold changes in gene expression using 2^{-ΔΔCt} method.

Statistical analysis

Data were analyzed using SPSS 11.5 (SPSS Inc, Chicago USA). The descriptive results are shown as frequency, mean and standard deviation. For training trial test by MWM, data were analyzed by one-way ANOVA and Bonferroni post hoc test for three recorded factors (escape latency, path length and the swimming speed) between groups separately by each day for five days. The trend for each test was analyzed by repeated measure analysis. The relative gene expression was compared between groups. Statistical analysis was performed by Kruskal Wallis test. The p-value less than 0.05 was considered statistically significant.

Results

Behavioral test

The results showed that in all study groups, the swimming distance and time for finding the hidden platform decreased during five days. The most prominent decreasing happened in the control group (normal rat); however, in the STZ induced rats' model, the average of total swimming distance and time in five days significantly increased in comparison to the control group (p=0.001). In herbal extract-treated group, the swimming distance and time had no significant difference in comparison to control group. Significant

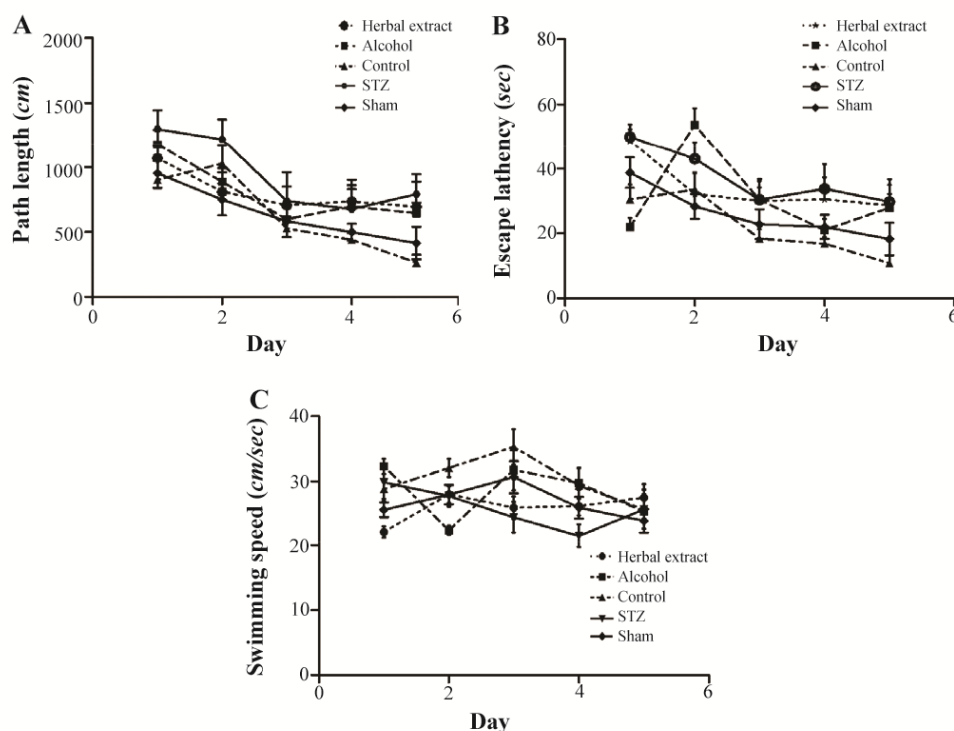


Figure 1. Trials performed in four continuous days. A) The mean value of the path length (swimming distance), B) the mean value of escape latency (time), and C) the mean value of swimming velocity (speed) in herbal extract, STZ, Alcohol, Sham and control groups.

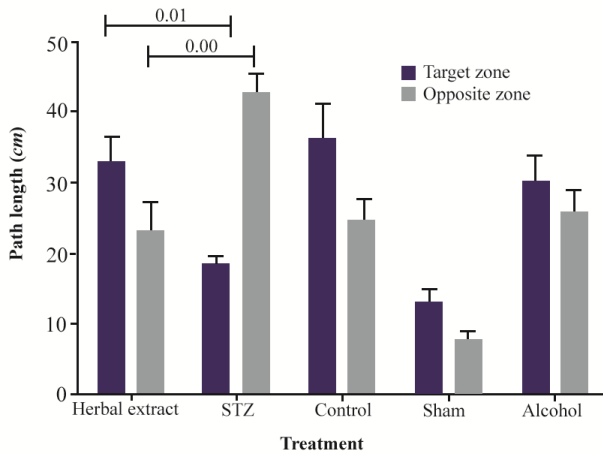


Figure 2. The mean value of total swimming distance spent in each quadrant by rat (percentage) in herbal extract, Alcohol, STZ, Sham and control.

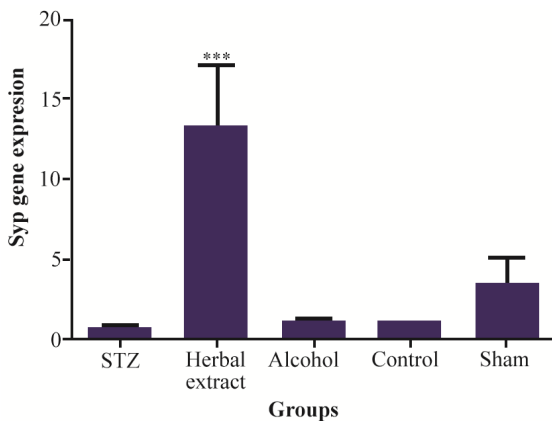


Figure 3. Comparison of synaptophysin gene expression level between the five study groups.

changes in spatial learning seen in the rat model group were improved in herbal-treated group. Alcohol and sham groups had no significant difference in swimming distance and time in the total days. The swimming speed was not statistically significant between groups (Figure 1).

The results of probe trial are shown in figure 2. The STZ group spent more time in opposite quadrant than in target zone but in herbal extract-treated group, each animal spent more time in the target zone than in the opposite quadrant, compared to the untreated group. And the difference between the treated and model group was significant.

Gene expressions

Syp, *Psen1* genes were found to have different expression in rat’s hippocampus after three weeks treatment with herbal extract when compared to other control groups, while the expression level of *Mapk3*, *Tnf-α*, *Mtap2* did not show any difference after 3 weeks of treatment. Statistically significant change, in expression level of *Syp* and *Psen1* were observed after three

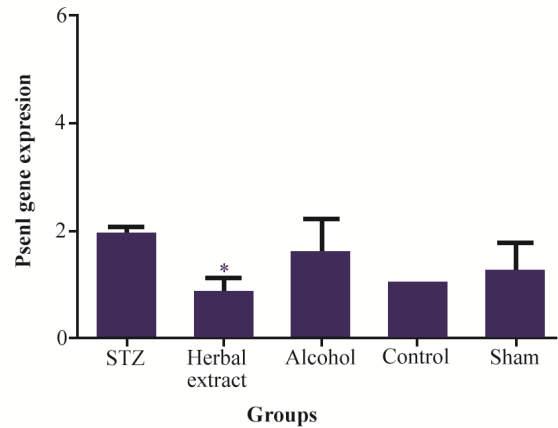


Figure 4. Comparison of *Psen1* gene expression level between the five study groups.

weeks of treatment with herbal extract, including a significant ($p=0.001$) twelve-fold increase in the expression level of *Syp* by herbal extract treatment (Figure 3). Meanwhile, the expression level of *Psen1* significantly decreased ($p=0.029$) one-half-fold (Figure 4). The decrease in expression of *Tnf-α* was most obvious but was not statistically significant. There was no significant difference of expression in the five genes between vehicle-treated, control and modeling groups after 3 weeks of treatment.

Discussion

Increasing evidence has supported an important role of gene expression in the initiation and progression of AD. Knowledge of the changes in the gene expression profile leads us to development of drug candidates that can reverse the changes in transcription and relieve the AD symptoms. In addition, change in gene expression probably occurs in the early stage of the disease and before the appearance of the pathological hallmarks. Therefore, drugs that can change the gene expression, can be potentially used for early treatment or prevention of the disease²⁷.

The loss of synaptic proteins, especially synaptophysin, is directly related to Alzheimer’s disease progression and cognitive decline. Evidence from mouse models showed that synaptophysin loss is most commonly seen in the hippocampus, than other brain regions and deficits in synapses are crucial to the development of the SAD^{28,29}. Interestingly, previous studies showed that SYP is a marker of SAD pathology even before formation of Aβ plaque³⁰. In this study, the influence of the herbal extract on *Syp* expression after treatment was demonstrated. In this investigation, a significant increase in *Syp* mRNA (induced with herbal extract treatment) was shown as well. According to previous studies, change in synaptophysin usually occurs before Aβ accumulation³¹. Therefore, this result implicates that up-regulation of *Syp* by herbal extract treatment, could be triggered as an early option for treatment of AD.

In recent years, several studies have shown an increase of *APP* mRNA levels in AD brains, which exacerbate amyloid- β deposition²⁷. *Psen1* as the catalytic subunit of γ -secretase has a known role in APP processing and A β formation. It was shown that significant decreases in *Psen1* mRNA were induced with herbal extract treatment. Therefore, herbal compounds such as *R. canina*, *T. vulgare* and *U. dioica*, which potentially could reduce the expression of *PSEN1*, may decrease the A β generation and would be an effective and novel drug for AD therapy²⁷.

It has been shown that the A β interacts in a synergistic way with cytokines to induce neuronal damage³². These events seem to be modulated by *TNF- α* which was up regulated in AD patient resulting in a sequence of events that lead to the generation of free radicals, oxidative damage and inflammation³³. A remarkable decrease in expression level of *Tnf- α* was observed after treatment with herbal extract. Interestingly, pharmacological inhibitors of *TNF- α* , like this herbal extract, would work as an immunomodulatory drug and improve learning and memory function. These protective effects are justified by reduction in the reactive oxygen species synthesis and synaptic disruption³⁴. Feedback loop of free radicals and A β finally leads to oxidation of protein and DNA, inhibition of ATP, neuronal damage and cognitive decline; however, the initial trigger is unknown since an age-dependent increase of oxidative stress has also been identified as an important factor leading to progression of Alzheimer's disease. Therefore, antioxidant and anti-inflammatory reagents like herbal extract of *R. canina*, *T. vulgare* and *U. dioica* can work as an anti-aging agent and prevent AD progression³⁵.

The Morris Water Maze system was applied in this study, which is used for the research in learning and memory and is directly related to the functions of the hippocampus²³. The results of the present study suggest that this herbal extract may shorten the total swimming time and the total swimming distance of the SAD rats. In probe test for testing memory ability of rats, herbal extract could improve their strategy for searching the target quadrant zone and decrease the swimming in opposite zone. Therefore, the above results could indicate that the mentioned component could improve the spatial learning and memory in SAD rat model.

Previous studies have shown that this herbal extract treatment would markedly enhance learning and memory in an aged mouse model²⁰. The present study shows that 3 weeks of treatment of SAD in the rat model with this compound (20 mg/kg/day) induces a significant influence in expression of *Syp* and *Psen1* and both of them are important for neuronal physiology, and pathogenesis of Alzheimer's disease. As considered in Morris Water Maze, significant changes that occurred in the rat model group were improved in herbal extract treated group. The changes seen in gene

expression level and behavioral test probably suggest that herbal extract of *R. canina*, *T. vulgare* and *U. dioica* has an anti-dementia property.

Acknowledgement

We would like to thank Rose PharMed Co. (Iran) for providing the total herbal extract. The study was supported by the University of Social Welfare and Rehabilitation Sciences, Tehran, Iran.

References

1. Diwu YC, Tian JZ, Shi J. Effects of Chinese herbal medicine Yinsiwei compound on spatial learning and memory ability and the ultrastructure of hippocampal neurons in a rat model of sporadic Alzheimer disease. *Zhong Xi Yi Jie He Xue Bao* 2011;9(2):209-215.
2. Mocerri VM, Kukull WA, Emanuel I, van Belle G, Starr JR, Schellenberg GD, et al. Using census data and birth certificates to reconstruct the early-life socioeconomic environment and the relation to the development of Alzheimer's disease. *Epidemiology* 2001;12(4):383-389.
3. Iqbal K, Grundke-Iqbal I. Metabolic/signal transduction hypothesis of Alzheimer's disease and other tauopathies. *Acta Neuropathol* 2005;109(1):25-31.
4. Tuppo EE, Arias HR. The role of inflammation in Alzheimer's disease. *Int J Biochem Cell Biol* 2005;37(2):289-305.
5. Yin Y, Liu Y, Huang L, Huang S, Zhuang J, Chen X, et al. Anti-apoptosis effect of astragaloside Iv on Alzheimer's disease rat model via enhancing the expression of Bcl-2 and Bcl-Xl. *Scand J Lab Anim Sci* 2010;37(2):75-82.
6. Chen Y, Tian Z, Liang Z, Sun S, Dai CL, Lee MH, et al. Brain gene expression of a sporadic (icv-STZ Mouse) and a familial mouse model (3xTg-AD mouse) of Alzheimer's disease. *PLoS One* 2012;7(12):e51432.
7. Callahan LM, Vaules WA, Coleman PD. Quantitative decrease in synaptophysin message expression and increase in cathepsin D message expression in Alzheimer disease neurons containing neurofibrillary tangles. *J Neuropathol Exp Neurol* 1999;58(3):275-287.
8. Colangelo V, Schurr J, Ball MJ, Pelaez RP, Bazan NG, Lukiw WJ. Gene expression profiling of 12633 genes in Alzheimer hippocampal CA1: transcription and neurotrophic factor down-regulation and up-regulation of apoptotic and pro-inflammatory signaling. *J Neurosci Res* 2002;70(3):462-473.
9. Rutten BP, Van der Kolk NM, Schafer S, van Zandvoort MA, Bayer TA, Steinbusch HW, et al. Age-related loss of synaptophysin immunoreactive presynaptic boutons within the hippocampus of APP751SL, PS1M146L, and APP751SL/PS1M146L transgenic mice. *Am J Pathol* 2005;167(1):161-173.
10. Ishibashi K, Tomiyama T, Nishitsuji K, Hara M, Mori H. Absence of synaptophysin near cortical neurons containing oligomer Abeta in Alzheimer's disease brain. *J Neurosci Res* 2006;84(3):632-636.
11. Theuns J, Van Broeckhoven C. Transcriptional regulation of Alzheimer's disease genes: implications for susceptibility. *Hum Mol Genet* 2000;9(16):2383-2394.

12. Chumakov I, Nabirotkin S, Cholet N, Milet A, Boucard A, Toulorge D, et al. Combining two repurposed drugs as a promising approach for Alzheimer's disease therapy. *Sci Rep* 2015;5:7608.
13. Iqbal K, Liu F, Gong CX, Alonso Adel C, Grundke-Iqbal I. Mechanisms of tau-induced neurodegeneration. *Acta Neuropathol* 2009;118(1):53-69.
14. Li R, Yang L, Lindholm K, Konishi Y, Yue X, Hampel H, et al. Tumor necrosis factor death receptor signaling cascade is required for amyloid-beta protein-induced neuron death. *J Neurosci* 2004;24(7):1760-1771.
15. Walsh DM, Selkoe DJ. Deciphering the molecular basis of memory failure in Alzheimer's disease. *Neuron* 2004;44(1):181-193.
16. Mohraz M, Khairandish P, Kazerooni PA, Davarpanah MA, Shahhosseiny MH, Mahdavian B, et al. A clinical trial on the efficacy of IMOD in AIDS patients. *Daru* 2009;17(4):277-284.
17. Mahmoodpoor A, Eslami K, Mojtahedzadeh M, Najafi A, Ahmadi A, Dehnadi-Moghadam A, et al. Examination of Setarud (IMOD™) in the management of patients with severe sepsis. *Daru* 2010;18(1):23-28.
18. Mohseni-Salehi-Monfared SS, Habibollahzadeh E, Sadeghi H, Baeeri M, Abdollahi M. Efficacy of Setarud (IMOD™), a novel electromagnetically-treated multiherbal compound, in mouse immunogenic type-1 diabetes. *Arch Med Sci* 2010;6(5):663-669.
19. Mohammadirad A, Khorram-Khorshid HR, Gharibdoost F, Abdollahi M. Setarud (IMOD™) as a multiherbal drug with promising benefits in animal and human studies: A comprehensive review of biochemical and cellular evidences. *Asian J Anim Vet Adv* 2011;6(12):1185-1192.
20. Ghanbari S, Yonessi M, Mohammadirad A, Gholami M, Baeeri M, Khorram-Khorshid HR, et al. Effects of IMOD™ and Angipars™ on mouse D-galactose-induced model of aging. *Daru* 2012;20(1):68.
21. Paxinos G, Watson C. The rat brain in stereotaxic coordinates. 6th ed. Australia: Academic press; 2004. 456 p.
22. Dhull DK, Jindal A, Dhull RK, Aggarwal S, Bhateja D, Padi SS. Neuroprotective effect of cyclooxygenase inhibitors in ICV-STZ induced sporadic Alzheimer's disease in rats. *J Mol Neurosci* 2012;46(1):223-235.
23. Zhou S, Yu G, Chi L, Zhu J, Zhang W, Zhang Y, et al. Neuroprotective effects of edaravone on cognitive deficit, oxidative stress and tau hyperphosphorylation induced by intracerebroventricular streptozotocin in rats. *Neurotoxicology* 2013;38:136-145.
24. National Research Council (US) Committee for the Update of the Guide for the Care and Use of Laboratory Animals. *Guide for the Care and Use of Laboratory Animals*. 8th ed. Washington (DC): National Academies Press (US); 2011. 246 p.
25. Silver N, Cotroneo E, Proctor G, Osailan S, Paterson KL, Carpenter GH. Selection of housekeeping genes for gene expression studies in the adult rat submandibular gland under normal, inflamed, atrophic and regenerative states. *BMC Mol Biol* 2008;9:64.
26. Moura AC, Lazzari VM, Agnes G, Almeida S, Giovenardi M, Veiga AB. [Transcriptional expression study in the central nervous system of rats: what gene should be used as internal control?] *Einstein (Sao Paulo)* 2014;12(3):336-341. English, Portuguese.
27. Chen XF, Zhang YW, Xu H, Bu G. Transcriptional regulation and its misregulation in Alzheimer's disease. *Mol Brain* 2013;6:44.
28. Selkoe DJ. Alzheimer's disease is a synaptic failure. *Science* 2002;298(5594):789-791.
29. Ingelsson M, Fukumoto H, Newell KL, Growdon JH, Hedley-Whyte ET, Frosch MP, et al. Early Abeta accumulation and progressive synaptic loss, gliosis, and tangle formation in AD brain. *Neurology* 2004;62(6):925-931.
30. Saganich MJ, Schroeder BE, Galvan V, Bredesen DE, Koo EH, Heinemann SF. Deficits in synaptic transmission and learning in amyloid precursor protein (APP) transgenic mice require C-terminal cleavage of APP. *J Neurosci* 2006;26(52):13428-13436.
31. Liu L, Orozco IJ, Planel E, Wen Y, Bretteville A, Krishnamurthy P, et al. A transgenic rat that develops Alzheimer's disease-like amyloid pathology, deficits in synaptic plasticity and cognitive impairment. *Neurobiol Dis* 2008;31(1):46-57.
32. Panahi N, Mahmoudian M, Mortazavi P, Hashjin GS. Effects of berberine on β -secretase activity in a rabbit model of Alzheimer's disease. *Arch Med Sci* 2013;9(1):146-150.
33. Sharma V, Deshmukh R. Tumor necrosis factor and Alzheimer's disease: A cause and consequence relationship. *Klinik Psikofarmakol Bülteni* 2012;22(1):86-97.
34. Medeiros R, Prediger RD, Passos GF, Pandolfo P, Duarte FS, Franco JL, et al. Connecting TNF-alpha signaling pathways to iNOS expression in a mouse model of Alzheimer's disease: relevance for the behavioral and synaptic deficits induced by amyloid beta protein. *J Neurosci* 2007;27(20):5394-5404.
35. Reddy PH. Amyloid precursor protein-mediated free radicals and oxidative damage: implications for the development and progression of Alzheimer's disease. *J Neurochem* 2006;96(1):1-13.